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Using ASAR for Analysis of Electrogenic and Human Gut Microbial Communities

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Abstract: In this paper we describe applications of our ASAR package to functional, taxonomic and pathways analysis of metagenomes and propose future plans and perspectives. To illustrate an analytical potential of ASAR, we discuss outcomes of several projects. The main focus is made on metabolic plasticity of electrochemically active microbial communities and a potential role of integrated symbiotic bacterial interactions; antipathogenic properties of BES, manifested in its capacity to remove some pathogens from waste streams; and medical applications of this technology. We present ASAR-based metagenome analysis of evolving bacterial community from distillery waste over period of 36 months in BES environment as an example. Application of ASAR to personalised analyses of gut microbiome (GM) and the data interpretation based on publicly available association studies are also discussed in this publication.

1 INTRODUCTION


For last years, we have been engaged in development of Bioelectrochemical Systems (BES)/Microbial Fuel Cell (MFC) technology for wastewater treatment. The BES/MFC applies complex interactions between microbial populations and electrodes to remove organics and to generate electricity (Gajda et al, 2018). By utilizing biological, chemical, engineering, and bioinformatics approaches, we seek to improve BES/MFC systems for better treatment efficiencies and electricity generation by understanding and building almost ideal microbial communities and developing cost-effective materials.


For better understanding of underlying biological processes, we have created pipeline for metagenome analysis. We have developed a new software, Advanced metagenomic Sequence Analysis in R (ASAR) (Orakov et al, 2017), which allows simultaneous analysis and visualization of taxonomic, functional, and pathways profiles of bacterial communities from the metagenome data. We have used ASAR to describe and improve


complex microbial communities and biofilms in BES/MFC. The ASAR package is available for researchers worldwide via GitHub. Statistical data analysis software has been integrated into the ASAR SA package, which includes capabilities to: plot Principle Coordinates Analysis (PCoA), perform distributed stochastic neighbour embedding (t-SNE), and to retrieve statistics estimated through the use of the pairwise permutational analysis of variance (PERMANOVA).

We have also developed flat FBA modelling software for community metabolomics studies and integrated it into the ASAR package (https://github.com/lptolik/asar_fba). Our ASAR DB (Orakov et al, 2017) includes more than 400 metagenomes and is the largest in the world electrogenic metagenomes proprietary database.

Via systematic analysis and recording of multiple samples we also found a range of species within the anode communities possessing the capacity for extracellular electron transfer, both via direct contact and electron shuttles and were able to detect differential distribution of bacterial groups on the

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carbon cloth and activated carbon granules of the anode surface (Kiseleva et al, 2015a). We have successfully applied our tools for identification and isolation of a new bacterial strain of *Thalassospira HJ* (Kiseleva et al, 2015b). Using computational pathway analysis and metabolic engineering, we have recently constructed a novel strain of electrogenic bacterium, *Arcobacter butzleri*, which allows single analyte (lactate and acetate) detection and can be used as a biosensor (Szydlowski et al, 2020).

Back, in 2015, we have pioneered with taxonomic and functional analysis of MFC communities from different geographical location. Taxonomic analysis showed that Proteobacteria, Bacteroidetes and Firmicutes were abundant in AD sludge from distinct climatic zones and constituted the dominant core of the MFC microbiomes. Functional analysis revealed species involved in degradation of organic compounds commonly present in food industry wastewaters (Kiseleva et al, 2015 a).

Accumulation of methanogenic Archaea was observed in the electrogenic biofilms, suggesting competition (Georg et al, 2019, Kaur et al, 2014a,b) or rather a symbiotic relationship between electrogenes and methanogens and a possibility for simultaneous electricity and biogas recovery from one integrated wastewater treatment system (Kiseleva et al, 2015a). Using our metagenomic approach we described the microbial diversity of the MFCs planktonic and anodic communities derived from two distinctively different inocula (Kiseleva et al, 2015a) to illustrate a consistency of the MFC community structure. Though two different archaea species, *M. barkeri* and *M. thermautotrophicus*, increased in the bacterial communities of swine and biogas waste inoculated MFCs, respectively (Vasieva et al, 2019), presence of Proteobacteria (mostly Deltaproteobacteria) phylum and eight *Geobacter* genus species as the predominant taxa in both MFCs anodic communities have been demonstrated.

Functional analyses of metagenomes from our lab scale experiments was sufficient to reveal metabolic changes between different species of the MFC dominant genus, *Geobacter*, suggesting that optimal nutrient utilization at the lowest electrode potential is achieved via genome rearrangements and a strong inter-strain selection, as well as adjustment of the characteristic syntrophic relationships. These observations show a certain degree of metabolic and genomic plasticity of electrochemically active bacteria and their communities in adaptation to adverse anodic and cathodic compartments (Szydlowski et al, 2019).

To study a functional adaptation of the electrogenic bacterial community in more detail we

have constructed a lab scale and enhanced pilot-scale reactor for nitrate removal in BES. Under applied potentials, BES biofilms were dominated by autotrophic denitrifying bacteria with a potential to accept electrons from the electrode (bacteria genera of *Galionella*, *Sideroxydans*, *Thiobacillus*) and heterotrophic bacteria that are capable to accept electrons from Fe²⁺ (bacteria genus of *Thauera*).

Bacterial community analysis based on shotgun sequencing from the 3-electrode reactors has confirmed metabolic adaptation of the electrochemically active bacterial communities to distinct anodic and cathodic environments. Functional analyses of metagenomes suggests that optimal nutrient utilization at the lowest electrode potential is achieved via genome rearrangements and a strong inter-strain selection. NADH-quinone oxidoreductase (*nuoB/C/G/L*, *nuoD*, *nuoH* genes) and NADH-ubiquinone oxidoreductase (*nad3*) genes show the strongest dependence on the applied potential and their abundancy evolves strongly over the period of the experiment (Szydlowski et al, 2019).

We have also demonstrated that such evolution was correlated with functional enrichment in metagenomes of genes encoding for particular motile (*motB*, *flgEF*, *fgrM*) factors and diminishing presence of genes encoding for virulent factors of several taxa, especially Enteronacteriaceae (*Shiegella*, *Vibrio*) and Firmicutes (*Enterococcus*, *Clostridia*, *Listeria*) (Vasieva et al, 2019, Ieropoulos et al, 2017).

The projects outlined here aim to demonstrate capabilities of the ASAR-based approach in taxonomic and functional analysis of bacterial communities and detection of the communities' adaptive processes at the genomic level.

2 EXAMPLE. METAGENOME ANALYSIS OF DISTILLERY WASTE MFC

2.1 Experimental Setup

This study was conducted at the Mizuho Shuzo Awamory Distillery Ltd., in Okinawa, Japan.

Metagenomics changes that occur during the initiation period of a 60 L serpentine-channelled MFC treating awamori distillery wastewater at a four-day retention time (0.54 L h⁻¹) at a constant 27°C in the laboratory were previously reported (Kiseleva et al, 2015a). Within the first 70 days of operation the MFC achieved 80% COD removal (2 kg COD m⁻³ d⁻¹). In this study we performed a three-year operation of a

multiple-MFC systems deployed at an awamori facility, operating at a similar flow rate but under ambient environmental conditions. Chemical analysis was carried out at the Okinawa Prefecture Environmental Science Centre.

- Staged installation of 3 tray modules with serpentine flow, air breathing cathode and composite activated carbon granule & glassy carbon cloth anode.
- Each tray module split into two 21L volumes, total module volume 50L
- Microbial Fuel cell cathode assembly, operated without catholyte. Pat No: US 8846220 B2
- Modules fed continuously from pilot site storage, distillery wastewater ratio increased to raise BOD loading
- pH adjusted manually in dosing tank to pH 6.8 – 7
- Modules underwent retrofitting several times during operation to increase feed inlets, improve function
- Nearly 3 years continuous operation

The observed steady increase in power production over time was consistent with previous reports showing that MFC power production correlates with the thickness of the anode-colonizing biofilm (Nevin et al, 2008). No evidence of longer-term operation leading to a state of biofilm “exhaustion” in which the performance of the electrogenic community declines (Kassongo et al, 2011) were found for this particular settings.

2.2 Bioinformatics Analysis

Whole genome sequences and 16S sequences were initially analysed using custom-developed pipeline, as described elsewhere (Orakov et al, 2017, Menze et al, 2016), as well as functional analysis using PALADIN (only applicable to WGS) (Westbroo et al, 2017). To study the selective enrichment of different samples, PCoA analysis was performed (Anderson, 2001), and plots were generated using EMPERor online tool. Compositional analysis of the community was performed in R version 1.4. (Vázquez-Baeza et al, 2013) with package compositions. Relative abundance was represented as composition with absolute geometry (rcomp). One-way ANOVA was conducted to verify significant difference in abundances of taxa between reactors (van den Boogaart et al, 2016). For visualization purposes, five most abundant genera in the inoculum and five most abundant genera at the final week sample were selected. PERMANOVA analysis was performed with Adonis function from vegan R package (Oksanen et al, 2018).

Functional (SEED/RAST) (Overbeek et al, 2005), taxonomic and KEGG Ontology (Kanehisa, 2000) annotations of reads tagged with md5 IDs from MG-RAST together with sample metadata, functional and taxonomic annotation hierarchy trees were generated and downloaded. Next, functional and taxonomic annotations were merged by identical md5's corresponding to unique read sequences. Then read counts were summed for reads with same function and taxon. Functional and taxonomic read annotations to lowest level were matched to lowest level annotations in their corresponding hierarchy trees to generate the whole phylogeny of each read. The result is the 3D dataset with axes of Functions, Taxonomy and Metagenome samples with hierarchy for former two.

Our post-annotation analysis and visualization tool ASAR (Orakov et al, 2017) uses data integration algorithm to merge taxonomic and functional data annotated at read level. The resulting 3D dataset with axes of Functions, Taxonomy and Metagenome samples is visualized via three heatmaps of each axis versus two others (F&T, F&M, T&M). Additionally, KEGG pathway enrichment sorting/heatmap and its map visualization are implemented. Advantages of the tool are:

- 1) Integrated functional and taxonomic analysis;
- 2) Comparative analysis of pathway enrichments;
- 3) KEGG pathway maps visualization.

The heatmaps show log abundance of reads annotated with selected function in particular taxon within particular community. On the KEGG map each functional box is split into sections corresponding to analysed bacterial communities. Relevant abundancy of each function in each community is colour coded from green (the lowest) to dark red (the highest proportion in the community).

2.3 Results

After 18 months of bacterial community evolution (Fig. 1) sequences associated with ‘Electron donating reactions’ and ‘Reverse electron flow’ functions have increased in abundancy only in the anodic metagenomes.

A following list of functional categories was associated with sequences which abundance increased in all MFC chambers (Fig.1):

- Arginine-Urea cycle
- Heat shock
- Riboflavin, FMN, FAD
- Fatty acids metabolic cluster
- Organic acids
- Electron accepting reactions, NAD and NADH
- Oxidative stress

- Central carbohydrate metabolism
- Fermentation
- One carbon metabolism

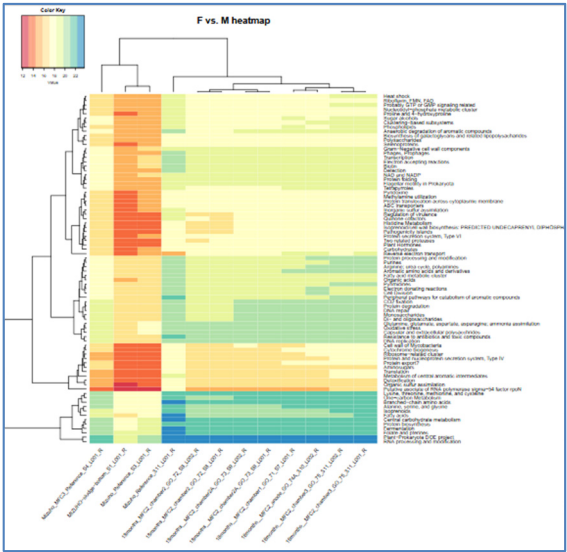


Figure 1: Relative abundances of reads (log scale) mapped to bacterial genomes in initial inoculum and in established MFC communities, generated via the ASAR taxa to samples function (level 3, max mapper (Orakov et al, 2017)). Log abundance is shown for reads annotated with selected functions in merged taxa within the metagenomes from anodic and planktonic communities from different MFC chambers after 18 months of initial bacterial community evolution.

After three years of cultivation several bacterial families became dominating and are specifically enriched in anodic metagenomes: Geobacteriaceae, Syntrophoaceae, Methanobacteriaceae. We also have noticed, that families of Clostridiaceae, Bacteroidaceae, Azonoxoaceae are increased mainly in chambers' metagenomes. At the level of genera the genomic presentation of following became obviously abundant in MFC (independently on a particular MFC's location): *Syntrophobacter*, *Syntrophus*, *Geobacter*, *Clostridium*, *Desulfovibrio*, *Bacteroides*, *Methanothermobacter*, *Thiobacillus*, *Dechloromonas*, *Metahnosphaera*, *Metahnobrevibacter*, *Metahnatrix*, *Pelobacter*, *Desulfobacillum*. *Syntrophobacter*, *Syntrophus*, *Geobacter* were stronger presented in anodic metagenomes. Abundances of genomic sequence presentation of the following genera were decreased in anodic metagenomes: *Dechloromonas*, *Clostridium*, *Bacteroides*, *Methanoregula*. Here the difference between the reference (s11) and the MFC metagenomes became very obvious. (Fig.2)

Using Canberra PCoA method (Fig. 3) we demonstrate a progressive change from the 3 month

community to 18 months community with 36 month community reversing to positions between 6 and 18 months data points. References data points are close to 6 months community ones or correspond to the stage before the 3 month community (close to the inoculum). Sludge reference data point is placed in between 18 and 36 months communities'.

Organic acids metabolism and fatty acid biosynthesis were among the most differentially expressed between the anodic samples from different stages of cultivation and in compare to the inoculum references. It is well presented in the corresponding KEGG maps. For instance, Fig. 4 presents the KEGG map for Butanoate metabolism for Geobacteriaceae family. One can see changes associated with particular evolutionary time points for 5 key functions.

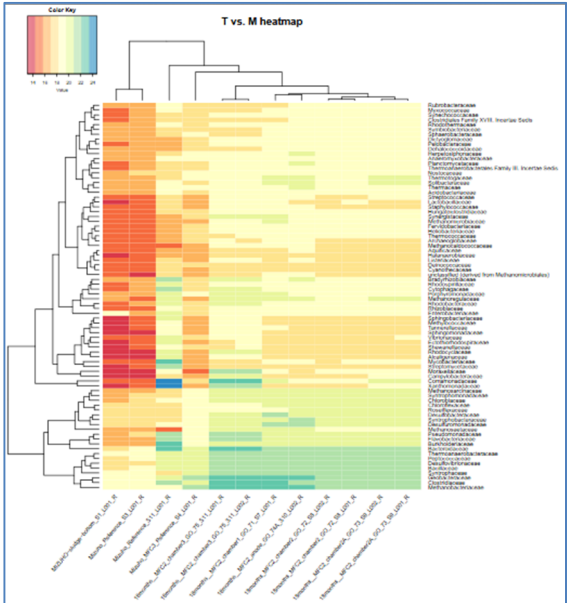


Figure 2: Relative abundances of reads (log scale) mapped to bacterial genomes in initial inoculum and in established MFC communities, generated via the ASAR taxa to samples function ('sum' option mapper (Orakov et al, 2017)). Log abundance is shown for merged reads annotated for selected taxa within the metagenomes from anodic and planktonic communities from different MFC chambers after 18 months of initial bacterial community (ref) evolution.

3 DISCUSSION

The results of the ASAR-based analysis of the metagenomes have been implemented in biotechnological projects that lead to optimisation of the MFC regimes and the bacterial strains, as well as generation of new hypothesis, which are awaiting

experimental validations. Our findings pointed to a potentially antipathogenic property of MFC and suggested that electrochemical metabolism may be utilized to suppress pathogenic bacteria without triggering a spread of antibiotics resistance (Vasieva et al, 2019, Ieropoulos et al, 2017). The highest loss among pathogenic genera was recorded for Enterobacteriaceae family (such as *Yersinia*, *Vibrio*, and *Shigella*). The abundance of virulent genes responsible for adhesion, secretion systems, invasion, and intracellular survival, as well as antibiotic resistance associated with Firmicutes and Actinobacteria phyla of Gram positive bacteria, also decreased in the MFC residential metagenomes (Vasieva et al, 2019).

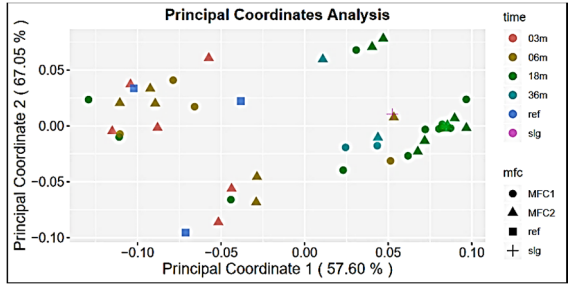


Figure 3: PCoA analysis for integrated taxonomic and functional data from metagenomes presenting 2 reference and 3, 6, 18, 36 months of bacterial community evolution.

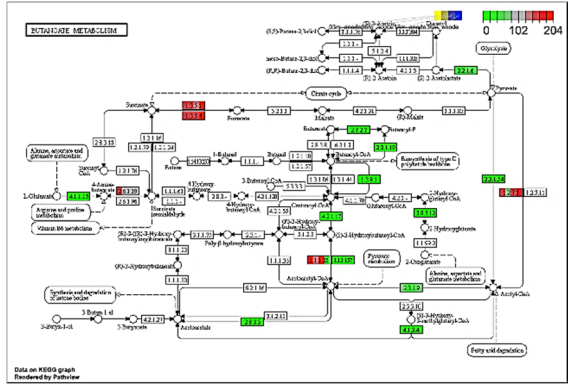


Figure 4: Butanoate metabolism. KEGG map for *Geobacteriaceae* reads enrichment in anodic communities. Different colours of parts of each block reflect levels of abundancies of the corresponding sequences in metagenomes from different community evolution time points (as indicated in the inserted legends).

Functional coupling and comparative genomics analysis have been applied to study functional associations of Enterococcal cAD1 sex pheromone precursor (P13268, *cad*) and its orthologs, known to be responsible for cell clumping, biofilm formation and conjugative plasmid transfer associated with

bacterial antibiotics resistance. Our analysis of genomic neighbourhood, motifs and phylogeny of *cad* shows that the cAD1 sex pheromone peptide release may depend on the precursor's redox properties, NADH and FMN-based redox metabolism (NADH oxidoreductase, fumarate reductase), and a FMN insertion chaperone, flavin trafficking facilitator ApbE (Q82Z24). We suggest a hypothetical model linking the NADH-driven and FMN-dependent redox metabolism and availability of soluble cofactors with *Enterococcus*, *Listeria*, *Oenococcus* and the relevant bacterial virulent properties during the operation of MFC for a purpose of waste waters and medical waste treatment (Vasieva and Goryanin, 2019). The novelty of the hypothesised association between sex pheromone release and the redox-related enzymatic function of the precursor lipoprotein suggests a new approach in prevention of antibiotic resistance spread via targeting sex pheromone processing chaperones or the cofactor availability.

We have validated our approach on personalised gut microbiome (GM) analysis and interpretation based on published association studies. We have applied the ASAR to a series of 10 sequenced GMs from individuals of different ethnical and geographical backgrounds, age and health groups. The differentially presented and detectable taxonomic and functional signatures in each GM metagenome were used to predict the hosts' characteristics via correlations established in published studies, and the predictions were validated by available individual-associated metadata. We have tested sensitivity of the routine annotation and data clustering pipeline to an individual and family-linked signatures in GM structure and functionalities, when applied to a limited number of varying samples. The number of samples was sufficient to demonstrate 2 main types of a GM composition, based on *Bacteroides* or *Prevotella* as the main abundant genera; limitation of a variety of taxa as a result of antibiotics application; clustering of family members' GM metagenomes, both in taxonomic and in functional space; individual signatures related to chronic diseases and pharmacological interventions; and elements of ethnicity related characteristics in the metagenomes (Vasieva et al, 2019c).

Cross-application of the approach to MFC and different from MFC's bacterial communities (such as Gut microbiomes) (Kaur et al, 2014, Ieropoulos et al, 2017, Vasieva et al, 2019) ensures more detailed validation of the developed analytical methods, and sets new standards for their improvement. With more bacteria genes becoming functionally annotated and

increasing understanding of metabolic logistics within an evolved bacterial community we are aiming to constantly refine our methods. However, it is time to learn principles of a microbiome adaptive evolution and the criteria and contrasts that we can use in the analysis, now and in the future.

4 CONCLUSIONS

We have shown that our computational pipeline and ASAR package could be successfully used in practical applications. We have analysed electrogenic and human microbial communities and produced novel data used for the software validation and prove of its capabilities. Original hypothesis were also generated which require further experimental confirmation.

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REFERENCES

- Anderson, M. J. 2001. A new method for non-parametric multivariate analysis of variance. *Austral Ecology*, 26: 32–46
- Gajda, I., Greenman, J., Ieropoulos, I.A., 2018. Recent advancements in real-world microbial fuel cell applications. *Curr Opin Electrochem*, 11:78-83, doi:10.1016/j.coelec.2018.09.006
- Georg, S., de Eguren Cordoba, I., Sleutels, T., Kuntke, P., terHeijne, A., Buisman, C. J. N., 2019. Competition of electrogens with methanogens for hydrogen in bioanodes. *Water Research*, 170:115292, doi: 10.1016/j.watres.2019.115292.
- Ieropoulos, I., Pasternak, G., Greenman, J., 2017. Urine disinfection and in situ pathogen killing using a microbial fuel cell cascade system. *PLoS One*, 12:1
- Kanehisa, M. and Goto, S., 2000. KEGG: Kyoto Encyclopedia of Genes and Genomes. *Nucleic Acids Res*, 28:27-30
- Kassongo, J., Togo, C. A., 2011. Performance improvement of whey-driven microbial fuel cells by acclimation of indigenous anodophilic microbes. *Afr J Biotechnol*, 10:7846–7852
- Kaur, A., Boghani, H. C., Michie, I., Dinsdale, R. M., Guwy, A. J., Premier, G. C., 2014. Inhibition of methane production in microbial fuel cells: operating strategies which select electrogens over methanogens. *Bioresource Technology*, 173:75-81, doi: 10.1016/j.biortech.2014.09.091
- Kiseleva, I., Garushyants, S.K., Ma, H., Simpson, D.J.W., Fedorovich, V., Goryanin, I., 2015. Taxonomic and functional metagenomic analysis of anodic communities in two pilot-scale microbial fuel cells treating different industrial wastewaters. *J Integr Bioinform*, 12(3):273
- Kiseleva, L., Garushyants, S. K., Briliute, J., Simpson, D.J.W., Cohen, M.F., Goryanin, I., 2015. Genome sequence of the electrogenic petroleum-degrading *Thalassospira* sp. strain HJ *Genome Announc*. 3 (3), e00483-15
- Menzel, P., Ng, K. L., Krogh, A., 2016. Fast and sensitive taxonomic classification for metagenomics with Kaiju. *Nature Communications*, 7: 11257
- Nevin, K. P., Richter, H., Covalla, S. F., Johnson, J. P., Woodard, T. L., Orloff, A. L., Jia, H., Zhang, M., Lovley, D. R., 2008. Power output and columbic efficiencies from biofilms of *Geobacter sulfurreducens* comparable to mixed community microbial fuel cells. *Environ Microbiol*, 10:2505–2514, doi:10.1111/j.1462-2920.2008.01675.x.
- Oksanen, J., Blanchet, F. G., Friendly, M., Kindt, R., 2018. *Vegan: Community Ecology Package*. R package, 2:4–6
- Orakov, A., Sakenova, N., Goryanin, I., Sorokin A., 2018. ASAR Database: An R Tool for Visual Analysis and Storage of Metagenomes in *Proceedings of the 11th International Joint Conference on Biomedical Engineering Systems and Technologies - Volume 4*. *Bioinformatics*, 196-200
- Orakov, A. N., Sakenova, N. K., Sorokin, A., Goryanin, I. I., 2017. ASAR: visual analysis of metagenomes in R. *Bioinformatics*, 34 (8): 1404-1405
- Overbeek, R., Begley, T., Butler, R. M., et al., 2005. The subsystems approach to genome annotation and its use in the project to annotate 1000 genomes. *Nucleic Acids Res*. 33(17):5691–5702, doi:10.1093/nar/gki866
- Szydlowski, L., Sorokin, Vasieva, O., Fedorovich, V., Goryanin, I., 2019. Evolutionary dynamics of microbial communities in bioelectrochemical systems. *bioRxiv*, 725580
- Szydlowski, L., et al, Goryanin, I., 2020. Novel strain of *Arcobacter* isolation and metabolic engineering (submitted, *Metabolic Engineering*)
- Van den Boogaart, K., Tolosana, R. and Bren, M., 2016. *Compositions: Compositional data analysis r-pack*, <http://sp.lyellcollection.org/> Vasieva, O., Sorokin, A., Szydlowski, L. and Goryanin, I., 2019. Do Microbial Fuel Cells have Antipathogenic Properties? *J Comput Sci Syst Biol*, 12:3, doi:10.4172/0974-7230.1000301 (a)
- Vasieva, O., Goryanin, I., 2019. Is there a function for a sex pheromone precursor? *Journal of Integrative Bioinformatics*, 16(4): 20190016, <https://doi.org/10.1515/jib-2019-0016> (b)
- Vasieva, O., Sorokin, A., Murzabae, M., Babiak, P., Goryanin, I., 2019. Study on analysis of personal gut microbiome. *Comput Sci Syst Biol*, 12(3):71-79 (c)

- Vázquez-Baeza, Y., M. Pirrung, A., Gonzalez, R., Knight, 2013. EMPeror: a tool for visualizing high-throughput microbial community data. *Gigascience*, 2:16
- Westbrook, A., Ramsdell, J., Schuelke, T., Normington, L., Bergeron, R. D., Thomas, W. K., and MacManes, M. D., 2017. PALADIN: protein alignment for functional profiling whole metagenome shotgun data. *Bioinformatics*, 33(10):1473–1478